

Supplementary Data of Manuscript number- AEM01948-23R1:

Title- Natural transformation specific DprA coordinate DNA double strand break repair pathways in heavily irradiated *D. radiodurans*

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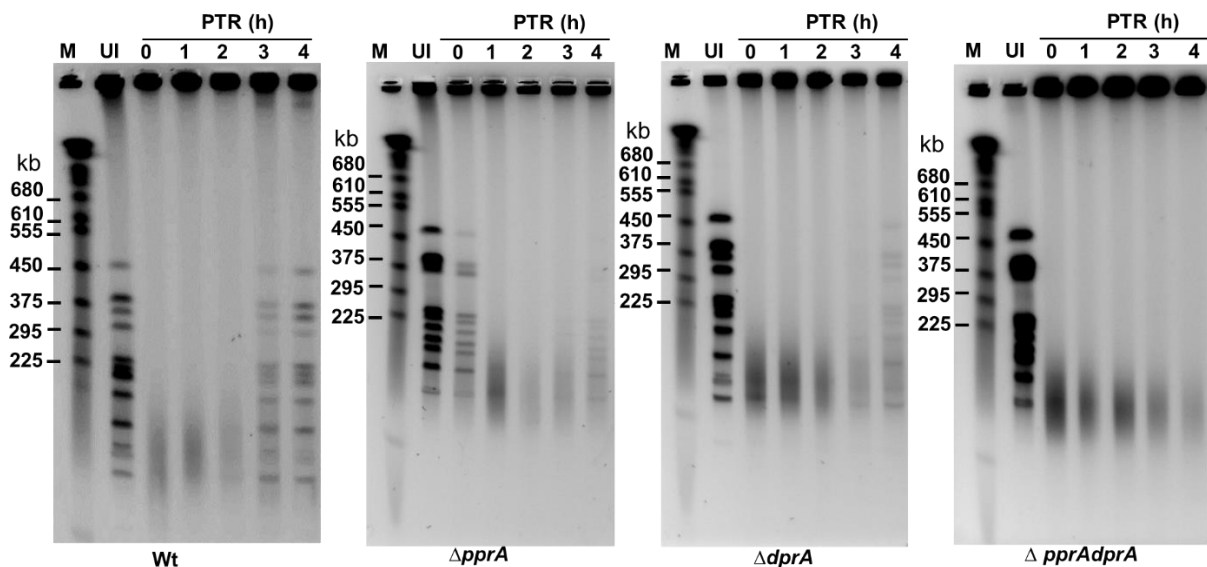


Fig. S1: The repair kinetics of DNA double-strand breaks (DSBs) in *D. radiodurans* wild-type and mutant strains upon treatment with MMC. The kinetics of DSB repair are shown for four strains, namely wild type (Wt), $\Delta pprA$, $\Delta dprA$, and $\Delta pprA \Delta dprA$ mutants. The evaluation of the repair kinetics was performed using pulsed-field gel electrophoresis (PFGE). The NotI-digested DNA from untreated cells (UI) and cells treated with 20 μ g/ml mitomycin C (MMC) for 30 min were allowed to recover in TGY medium for 4 hours post-treatment (PTR). Cell samples collected at different time points were analyzed on PFGE to assess the kinetics of DSB repair. *S. cerevisiae* molecular mass standards (lane-M).

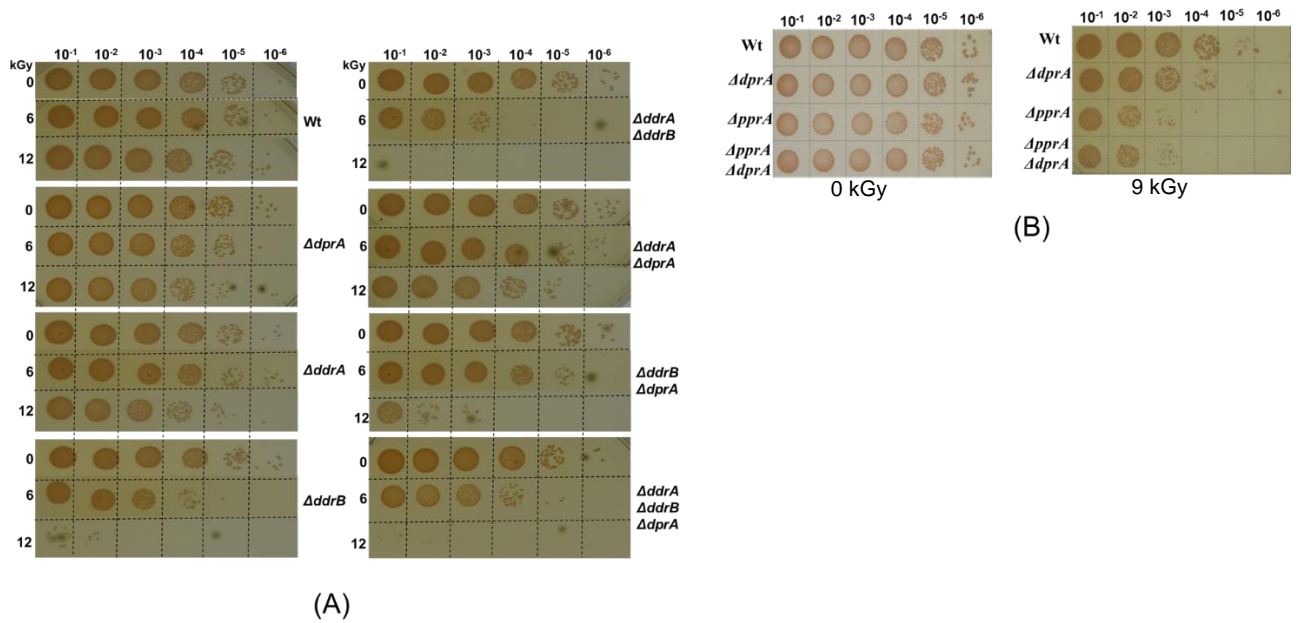


Fig. S2: Gamma radiation survival of various mutants of *D. radiodurans*.

(A) Exponential growth phase cells of wild type and mutants including $\Delta dprA$, $\Delta ddrA$, $\Delta ddrB$, $\Delta ddrA \Delta ddrB$, $\Delta ddrA \Delta dprA$, $\Delta ddrB \Delta dprA$, and $\Delta ddrA \Delta ddrB \Delta dprA$ were exposed to gamma radiation doses ranging from 0 to 12 kGy. After dilution, aliquots were spotted on TGY medium and incubated at 32°C for 48 hours.

(B) Wild type and mutants including $\Delta dprA$, $\Delta pprA$ and $\Delta pprA \Delta dprA$ were exposed to 9 kGy gamma radiation doses. After dilution, aliquots were spotted on TGY medium and incubated at 32°C for 48 hours.

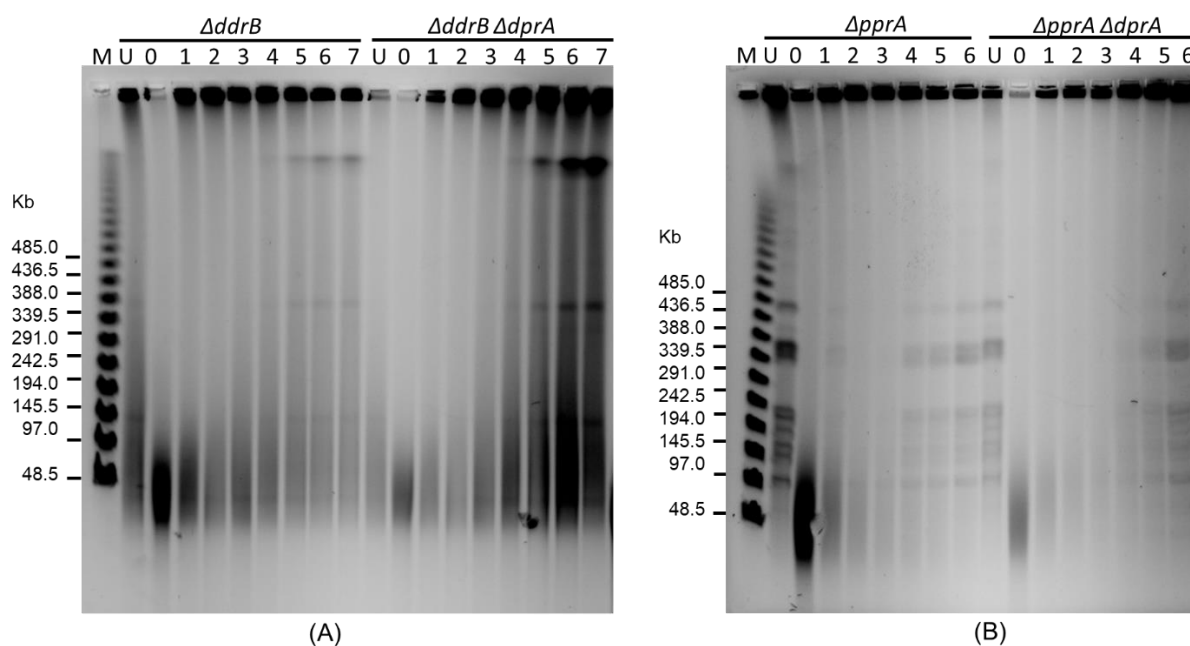


Fig. S3: The DNA double-strand breaks (DSBs) repair kinetics of *D. radiodurans* wild-type and its mutants.

PFGE was employed to assess the repair kinetics of DSBs. Panel (A) presents the repair kinetics of DSBs in $\Delta ddrB$ and $\Delta ddrB \Delta dprA$ mutants, while panel (B) displays the repair kinetics of DSBs in $\Delta pprA$ and $\Delta pprA \Delta dprA$ mutants. The NotI-digested DNA samples obtained from unirradiated cells (U) and cells irradiated with 6 kGy at different post-irradiation time points (PIR) were visualized on the gel immediately after irradiation (0) and at specific incubation times (in hours). Lambda PFG molecular mass standards were loaded in lane-M for size reference. The data suggests that the $\Delta ddrB \Delta dprA$ mutant exhibits relatively improved DSB repair compared to the $\Delta ddrB$ mutant, while the $\Delta pprA \Delta dprA$ mutant shows weaker DSB repair compared to the $\Delta pprA$ mutant.

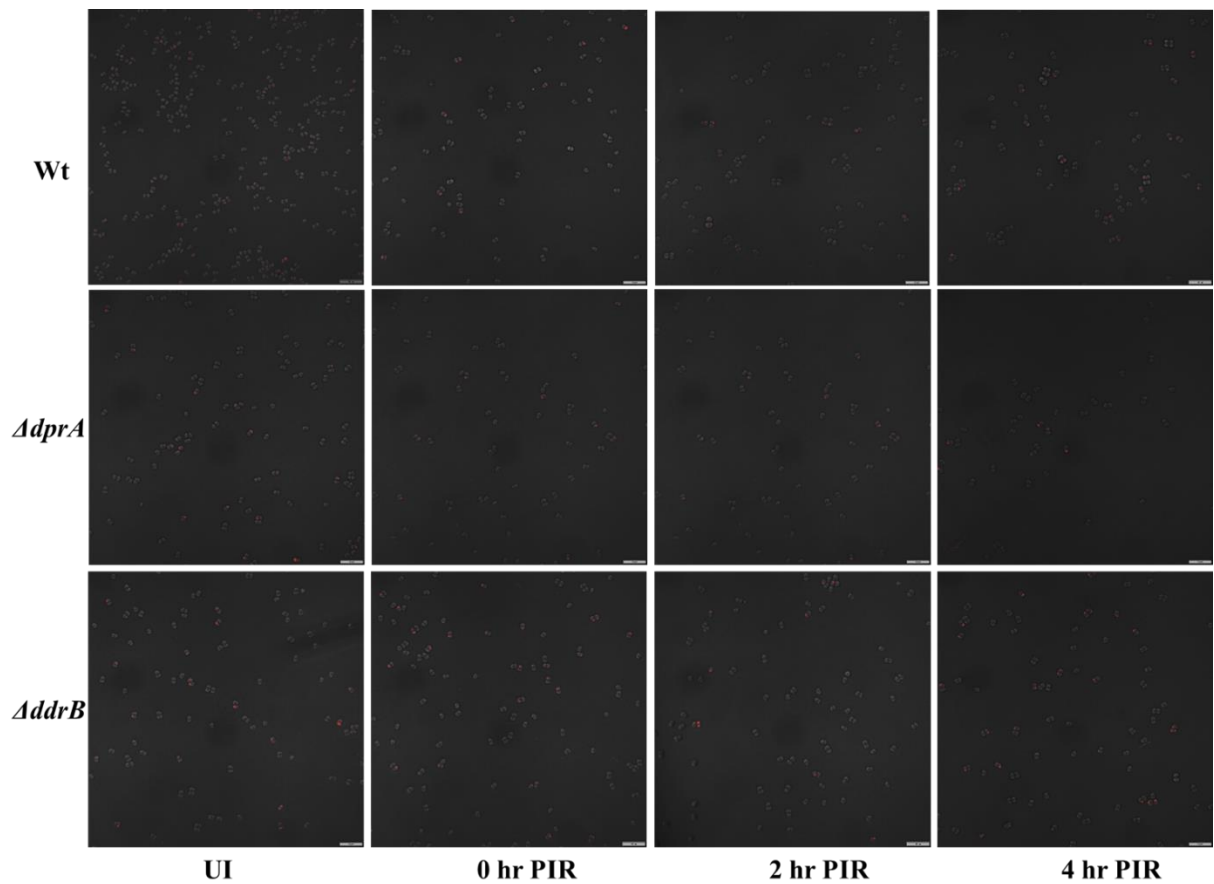


Fig. S4 RecA^{RFP} foci formation in wild type, *ΔdprA* and *ΔddrB* mutants.

For fixed-cell imaging, *D. radiodurans* cells expressing pRADrecA-RFP were fixed and observed using Olympus FV3000 confocal microscope, specifically. These cells were examined in the RFP channel (561 nm) and under bright field (DIC) to determine the localization of RecA^{RFP}. Image analysis was performed using automated cellSens software. The scale bar indicates a length of 10 μm.

Table S1: Statistical analysis of RecA^{RFP} foci formation in wild-type, *ΔddrB*, and *ΔdprA* mutants

Tukey's multiple comparisons test	Mean Diff.	95.00% CI of diff.	Significant?	Summary	Adjusted P Value
Wt vs. <i>ΔddrB</i>	51.72	27.12 to 76.32	Yes	**	0.0064
Wt vs. <i>ΔdprA</i>	52.17	27.58 to 76.77	Yes	**	0.0062
<i>ΔddrB</i> vs. <i>ΔdprA</i>	0.4550	-24.14 to 25.05	No	ns	0.9967

Table S2: Bacterial strains, plasmids, and primers used in this study

Bacterial strains	Organism	Genotype	Source
<i>D. radiodurans</i> R1	<i>D. radiodurans</i>	Wild type strain ATCC13939	Lab stock
<i>E. coli</i> Novablue	<i>E. coli</i>	<i>endA1 hsdR17(r_{K12}⁻ m_{K12}⁺) supE44 thi-1 recA1 gyrA96 relA1 lacF''[proA⁺B⁺ lacI^q ZΔM15::Tn10] (Tet^R)</i>	Lab stock
$\Delta pprA$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>pprA</i> Ω <i>chl</i>	Prof. Issay Narumi lab
$\Delta ddrA$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrA</i> Ω <i>chl</i>	Prof. Pascale Servant lab
$\Delta ddrB$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrB</i> Ω <i>kan</i>	Prof. Pascale Servant lab
$\Delta ddrA \Delta ddrB$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrA</i> Ω <i>chl</i> , <i>ddrB</i> Ω <i>kan</i>	Prof. Pascale Servant lab
$\Delta pprA \Delta dprA$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>pprA</i> Ω <i>chl</i> , <i>dprA</i> Ω <i>spec</i>	This work
$\Delta dprA$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>dprA</i> Ω <i>spec</i>	This work
$\Delta ddrA \Delta dprA$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrA</i> Ω <i>chl</i> , <i>dprA</i> Ω <i>spec</i>	This work
$\Delta ddrB \Delta dprA$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrB</i> Ω <i>kan</i> , <i>dprA</i> Ω <i>spec</i>	This work
$\Delta ddrA \Delta ddrB \Delta dprA$	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrA</i> Ω <i>chl</i> , <i>ddrB</i> Ω <i>kan</i> , <i>dprA</i> Ω <i>spec</i>	This work
<i>D. radiodurans</i> (<i>thy</i> ⁻)	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>thy</i> ⁻	This work
$\Delta dprA$ (<i>thy</i> ⁻)	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>dprA</i> Ω <i>spec</i> , <i>thy</i> ⁻	This work
$\Delta pprA$ (<i>thy</i> ⁻)	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>pprA</i> Ω <i>chl</i> , <i>thy</i> ⁻	This work
$\Delta pprA \Delta dprA$ (<i>thy</i> ⁻)	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>pprA</i> Ω <i>chl</i> , <i>dprA</i> Ω <i>spec</i> , <i>thy</i> ⁻	This work
$\Delta ddrB$ (<i>thy</i> ⁻)	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrB</i> Ω <i>kan</i> , <i>thy</i> ⁻	This work
$\Delta ddrB \Delta dprA$ (<i>thy</i> ⁻)	<i>D. radiodurans</i>	<i>D. radiodurans</i> R1 <i>ddrB</i> Ω <i>kan</i> , <i>dprA</i> Ω <i>spec</i> , <i>thy</i> ⁻	This work
Plasmids:			
Names	Characteristics and Source		lab stock
pRADgro	pRAD1 carrying 261bp <i>Bgl</i> II- <i>Xba</i> I fragment of promoter (Pgro) from <i>D. radiodurans</i>		lab stock
pDSRED-recA	pDSRED carrying <i>drrecA</i> at <i>Bam</i> HI and <i>Kpn</i> I		[64]
pRADrecA-RFP	pRAD carrying <i>drrecA</i> at <i>Apa</i> I and <i>Eco</i> RV		[64]
pNOSdprA	pNOS suicidal vector carrying upstream and downstream DNA fragment of <i>dprA</i> gene (<i>dr_0120</i>)		This work
Primer details			
Sl. No.	Name of primer and Sequence (5' to 3')		Purpose
1.	<i>dprA</i> -UF (ATA GGT ACC TAA GCG CCT ATC AAG CCC TC)		$\Delta dprA$ mutant generation
2.	<i>dprA</i> -UR (AAA GGG ATG CCG CCG CAG GTT TTC GAT)		
3.	<i>dprA</i> -DF (ATA GGA TCC GAA CTG AAC AAG GCG GCA GA)		
4.	<i>dprA</i> -DR (TAG TCT AGA ACG CGG CAA CAG AGA GAA GTC)		